

# Enhancement of Gap Junctional Communication and Connexin43 Expression by Thyroid Hormones

Anke Stock, Helmut Sies and Wilhelm Stahl\*

Institut für Physiologische Chemie I, Heinrich-Heine-Universität Düsseldorf, Postfach 101007, D-40001 Düsseldorf, Germany.

**ABSTRACT.** Cells in tissues coordinate their activity by sharing ions, second messengers, and small metabolites through clusters of intercellular channels called gap junctions. The thyroid hormones 3,3′,5-triiodo-L-thyronine (T<sub>3</sub>) and L-thyroxine (T<sub>4</sub>) are capable of modulating gap junctional communication (GJC) as are 1,25-dihydroxyvitamin D<sub>3</sub>, retinoic acid, and other nuclear receptor ligands. T<sub>3</sub> and T<sub>4</sub> were found to stimulate GJC in WB-F344 rat liver epithelial cells dose-dependently at concentrations between 1 nM and 0.1 μM, assayed by the dye transfer method using Lucifer Yellow CH. The stimulation of cell-cell communication was preceded by an increase in connexin43 mRNA levels and was accompanied by an accumulation of connexin43 protein measurable 2 days after incubation with these compounds. These observations establish a novel role of thyroid hormones in the regulation of gap junctional intercellular communication via connexin43 gene expression. BIOCHEM PHARMACOL 55;4:475–479, 1998. © 1998 Elsevier Science Inc.

**KEY WORDS.** intercellular communication; thyroid hormones; connexin43; up-regulation; epithelia; WB-F344

Thyroid, retinoid and steroid hormones play an important role in development, differentiation, and physiological responses to diverse stimuli. These molecules bind to specific intracellular receptors which belong to a superfamily of regulatory proteins activating the expression of gene networks [1]. The thyroid hormones 3,3',5-triiodo-L-thyronine  $(T_3)^{\dagger}$  and L-thyroxine  $(T_4)$  are ligands of the nuclear thyroid receptor (TR) [2]. Direct interaction of T<sub>3</sub>-liganded TRs with DNA sequences in the promoter of target genes, referred to as thyroid response elements, are commonly assumed to elicit a rapid transcriptional response even after 20 min [3]. The mRNA of rat growth hormone in rat pituitary tumour cells GH1 [4] and glucokinase mRNA in rat liver tissue [5] are both induced 3-fold within 2 hr after incubation with T<sub>3</sub> or T<sub>4</sub>. Phosphoenolpyruvate carboxykinase mRNA increases 2-fold within 3 hr [6].

Ligands for nuclear receptors such as retinoic acid [7] and 1,25-dihydroxyvitamin D<sub>3</sub> [8, 9] are known to induce gap junctional communication (GJC). Interactions between nuclear receptors of this superfamily [10, 11] might play a role in the control of GJC in different tissues. Connexinbased intercellular channels are selectively permeable to

many small molecules. Thus, GJC may influence a wide variety of cellular activities, including the regulation of growth, differentiation, and developmental signaling [12]. Pathological implications of raised GJC are, e.g., cardiac arrhythmias and heart failure [13, 14]. Similar effects have been described in context with hyperthyroidism [15]. Thus, aspects of the pathophysiology of hyperthyroidism could be associated with modulation of GJC. In the present study, we describe the influence of thyroid hormones T<sub>3</sub> and T<sub>4</sub> on GJC and their effects on connexin43 gene expression.

## MATERIALS AND METHODS Chemicals

Lucifer Yellow CH was purchased from Sigma, 3,3',5-triiodo-L-thyronine ( $T_3$ ) and L-thyroxine ( $T_4$ ) from Aldrich, morpholinopropane sulfonic acid from Serva and the oligolabeling kit from Pharmacia. All other chemicals were obtained from Merck.

#### Cells and Culture Conditions

The WB-F344 rat liver epithelial cells were a kind gift from Dr. I.A. Cotgreave and Dr. L. Wärngard, Institute of Environmental Medicine, Karolinska Institute (Stockholm, Sweden). The human embryonal skin fibroblast cell line HFFF2 was obtained from ECACC. The cells were cultured in Dulbecco's modification of Eagle's minimal essential medium (DMEM; Gibco BRL) supplemented with fetal calf

<sup>\*</sup> Corresponding author: Dr. W. Stahl, Institut für Physiologische Chemie I, Heinrich-Heine-Universität Düsseldorf, P.O. Box 101007, D-40001 Düsseldorf, Germany. Tel. +49 211 81 12711; FAX +49 211 81 13029; E-mail: wilhelm.stahl@uni-duesseldorf.de

<sup>†</sup>Abbreviations: T<sub>3</sub>, 3,3',5-triiodo-L-thyronine; T<sub>4</sub>, L-thyroxine; TR, thyroid hormone receptor; GJC, gap junctional communication; FCS, fetal calf serum; RA, retinoic acid.

Received 12 May 1997; accepted 5 August 1997.

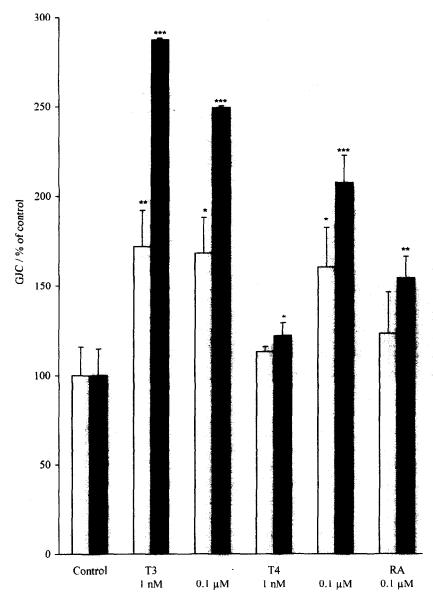


FIG. 1. Stimulation of gap junctional communication (GJC) by 3,3',5-trilodo-L-thyronine  $(T_3)$ , L-thyroxine  $(T_4)$  and retinoic acid (RA) in WB-F344 cells. Confluent cells were treated as described in "Materials and Methods." At day 1 (open bars) and 2 (solid bars), GJC was determined by the microinjection/dye transfer assay. Basal level of communicating cells was 23.8  $\pm$  3.5. Mean values of 10 individual injections were determined in four independent experiments and results standardized (n = 4). \*P < 0.05, \*\*P < 0.005, and \*\*\*P < 0.001.

serum 10% v/v (FCS; Greiner) and 2 mM glutamine (growth medium). Cells were grown on plastic dishes (Falcon) in a humidified incubator under an atmosphere of 5% CO<sub>2</sub> in air.

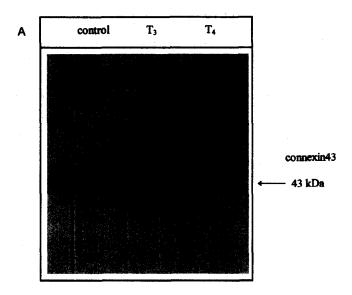
### Gap Junctional Communication

Gap junctional intercellular communication was assessed by the efficacy of diffusion of Lucifer Yellow CH (10% in 0.33 M LiCl w/v) from a single microinjected cell to its neighboring cells [16]. Confluent cells on 35 mm plates were incubated in growth medium without FCS for 24 hr, and then retinoic acid (RA) at 0.1 µM, 3,3',5-triiodo-L-thyronine (T<sub>3</sub>) or L-thyroxine (T<sub>4</sub>) at 0.1 µM–0.1 pM dissolved in phosphate buffer (pH 8) was added. After one and two days of incubation, microinjection was performed using microinjector 5242 and micromanipulator 5170 (Eppendorf). Dye-coupled cells were counted 5 min after injection. Mean values of 10 individual injections in cells

were determined. The mean values of 4 independent experiments are given as percentage of control. Differences in GJC between controls and treated cells were calculated for significance using Student's t-test. Data obtained with controls were set to 100%.

#### Connexin43 Protein

Confluent cells grown on 60 mm plates were incubated in growth medium without FCS for 24 hr, and  $T_3$  or  $T_4$  dissolved in phosphate buffer (pH 8) was added. Twenty-five  $\mu$ g cellular protein, isolated after a two-day incubation with the test compounds (0.1  $\mu$ M), was separated by SDS-PAGE and electrotransferred onto a nylon membrane. After incubation with mouse monoclonal antibodies generated to a peptide containing amino acids 252–270 of rat connexint3 (Affinity) at a dilution of 1:2000 in blocking buffer (ICN), detection was with the AURORA Western blotting kit (ICN).



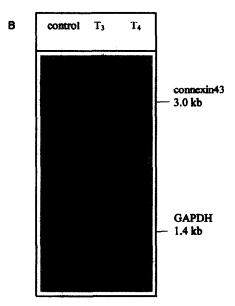


FIG. 2. Induction of connexin43 protein (A) and mRNA (B) by 3,3',5-triiodo-L-thyronine ( $T_3$ ) and L-thyroxine ( $T_4$ ) in WB-F344 rat liver epithelial cells. (A) Cells were incubated for 2 days with the thyroid hormones at 0.1  $\mu$ M, and Western blot analysis was performed with 25  $\mu$ g cellular protein in each lane. (B) Cells were incubated for three hr with the thyroid hormones at 0.1  $\mu$ M, and Northern blot analysis was performed. The figure shows representative results of experiments repeated at least three times each.

#### Connexin43 mRNA

Confluent cells grown on 100 mm plates were incubated in growth medium without FCS for 24 hr. Total RNA was isolated according to the acid phenol method [17]. Equal amounts of RNA (25  $\mu$ g), isolated after a 3, 6, and 9 hr incubation of confluent cells with T<sub>3</sub> or T<sub>4</sub> at 0.1  $\mu$ M, were separated by agarose gel electrophoresis and transferred onto a nylon membrane. Connexin43 mRNA was measured by hybridization with a radioactively labeled 0.7 kb AccI fragment of mouse connexin43, kindly provided by Dr. K. Willecke (Bonn, Germany) [18]. Glyceraldehydephos-

phate dehydrogenase (GAPDH), used for standardization, was measured using a radioactively labeled 1.3 kb *EcoRI* fragment of the plasmid pRLCGAP, kindly provided by Dr. R. Wu (Ithaca, NY, USA) [19]. Densitometric analysis was performed using a gel scanner with GSXL software (Pharmacia). Contents of connexin43 mRNA related to GAPDH mRNA are expressed as a percentage of solvent controls; mean values  $\pm$  SEM of three independent experiments are given.

#### RESULTS AND DISCUSSION

The present study demonstrates a novel role of thyroid hormones 3,3',5-triiodo-L-thyronine ( $T_3$ ) and L-thyroxine  $(T_4)$  in the modulation of gap junctional communication (GJC). Both compounds are capable of inducing GJC, as measured in the dye-transfer assay (Fig. 1), in WB-F344 rat liver epithelial cells which express connexin43 [20]; no morphological changes in the cells were apparent. After incubation of WB-F344 cells with 0.1 µM or 1 nM of T<sub>3</sub> (physiological concentration in human serum: 1.5 nM T<sub>3</sub>), GJC increased ca. 1.7-fold at day 1 over solvent control. A further increase to approximately 2.7-fold was observed at day 2. At lower concentrations, the effects of T<sub>3</sub> on GJC were less pronounced; no induction was observed at 0.1 pM (data not shown). The influence of  $T_4$  (physiological concentration in human serum: 0.1 µM T<sub>4</sub>) on GJC was less than that of T<sub>3</sub>. Hardly any induction was observed with 1 nM T<sub>4</sub> on days 1 and 2, while a 2-fold increase was detected with 0.1 µM T<sub>4</sub> at days 1 and 2. Inductory effects of T<sub>3</sub> were also found in the embryonal human skin fibroblasts HFFF2. There was a 1.8-fold increase in GJC after a 6-day incubation with 0.1 µM T<sub>3</sub>, but no significant increase was observed after incubation with T<sub>4</sub> (data not shown). The response of WB-F344-cells to treatment with retinoic acid (0.1  $\mu$ M), a compound which stimulates GJC in fibroblasts and various other cell lines, was lower, only 1.5-fold at day 2 (Fig. 1). With retinoic acid, increases were observed after 1 hr in the human amniotic cell line FL [21], but only after several days in murine fibroblasts C3H/10T1/2 [22, 23] or human dermal fibroblasts [24].

The up-regulation of GJC after incubation with T<sub>3</sub> and T<sub>4</sub> is associated with a 4- to 5-fold increase in connexin43 protein as shown by Western blot analysis after two days of incubation with both compounds (Fig. 2A). The two bands corresponding to 45 kDa and 47 kDa represent phosphorylated connexin43 [25]. Rises in connexin43 protein levels are preceded by increases in connexin43 mRNA levels. The levels of connexin43 mRNA were already elevated ca. 1.6to 2-fold at 3 hr (Fig. 2B). The effect of T<sub>3</sub> or T<sub>4</sub> on mRNA levels was similar and remained constant over a period of 9 hr (Table 1). No increase in connexin43 mRNA was observed with all-trans retinoic acid within this period (data not shown). In the human embryonal skin fibroblast cell line HFFF2, similar effects of T<sub>3</sub> on connexin43 mRNA levels were observed. After 5 to 9 hr of incubation, a 1.6-2 fold increase in mRNA levels was detected.

TABLE 1. Induction of connexin43 mRNA by 3,3'-5-triiodo-L-thyronine and L-thyroxine at 0.1 µM concentration in WB-F344 rat liver epithelial cells

Incubation time (hr)	Connexin43 mRNA Level (fold increase)	
	3,3',5-triiodo-L- thyronine	L- thyroxine
3	$1.8 \pm 0.1$	$1.8 \pm 0.2$
5	$1.4 \pm 0.03$	$1.6 \pm 0.5$
9	$1.7 \pm 0.4$	$1.8 \pm 0.6$

Data are given as means ± SEM for three independent experiments. Similar results were obtained with the human fibroblasts HFFF2.

The mechanism underlying stimulation of GJC seems to be different in different cell types. An increased level of connexin43 mRNA and protein has been described after treatment with all-trans-retinoic acid in murine C3H/ 10T1/2 fibroblasts [26], while in the rat liver epithelial cell line IAR203, retinoic acid enhances connexin43 protein levels without increasing mRNA [27]. Accumulated mRNA might result either from elevated transcription rates or from stabilization of mRNA. The stimulation of GJC by all-trans retinoic acid in mouse F9 teratocarcinoma cells has been ascribed to stabilization of connexin43 mRNA [28]. The promoter region of the rat connexin43 gene [29] contains a direct repeat (DR3) consensus sequence at -480 bp to -464 bp, a possible binding site, for example, for nuclear receptor heterodimers containing TR [30]. Thus, the relatively rapid transcriptional response of T<sub>3</sub> and T<sub>4</sub> may be due to direct regulation of the connexin43 gene.

Hyperthyroidism causes, among other adverse effects, atrial arrhythmias and congestive heart failure [15]. In addition, raised GJC has been related to pathological conditions such as cardiac arrhythmias and heart failure [13, 14]. Here we show for the first time that thyroid hormones influence GJC in vitro. It might be speculated that this effect is involved in the biochemical mechanism underlying the pathological effects of hyperthyroidism.

We thank Dr. A. Clairmont and Dr. C. Carlberg for helpful discussions. This work was supported by the Deutsche Forschungsgemeinschaft (Si 255/10-1), by the National Foundation for Cancer Research (Bethesda, MD) and the Bundesministerium für Bildung, Wissenschaft, Forschung und Technologie (Bonn, Germany).

#### References

- 1. Evans RM, The steroid and thyroid hormone receptor super-
- family. Science 240: 889-895, 1988.

  2. Bolger MB and Jorgensen EC, Molecular interactions between thyroid hormone analogs and the rat liver nuclear receptor. J Biol Chem 255: 10271–10278, 1980.

  3. Jump DB, Naryan P, Towle H and Oppenheimer JH, Rapid
- effects of triiodothyronine on hepatic gene expression. J Biol Chem 259: 2789-2797, 1984.
- 4. Yaffe BM and Samuels HH, Hormonal regulation of the growth hormone gene. J Biol Chem 259: 6284-6291, 1984.
- 5. Höppner W, Süssmuth W and Seitz HJ, Cooperative effect of

- thyroid hormones on glucokinase gene transcription in rat liver. J Biol Chem 264: 20643-20647, 1989.
- Höppner W, Süssmuth W and Seitz HJ, Cooperative effect of thyroid and glucocorticoid hormones on the induction of hepatic phosphoenolpyruvate carboxykinase in vivo and in cultured hepatocytes. Eur I Biochem 159: 399-405, 1986.
- 7. Mehta PP, Bertram JS and Loewenstein WR, The actions of retinoids on cellular growth correlate with their actions on gap junctional communication. J Cell Biol 108: 1053-1065, 1989.
- Stahl W, Nicolai S, Hanusch M and Sies H, Vitamin D influences gap junctional communication in C3H/10T1/2 murine fibroblast cells. FEBS Lett 352: 1-3, 1994.
- Clairmont A, Tessmann D, Stock A, Nicolai S, Stahl W and Sies H, Induction of gap junctional intercellular communication by vitamin D in human skin fibroblasts is dependent on the nuclear vitamin D receptor. Carcinogenesis 17: 1389-1391, 1996.
- 10. Kliewer SA, Umesono K, Mangelsdorf DJ and Evans RM, Retinoid X receptor interacts with nuclear receptors in retinoic acid, thyroid hormone and vitamin D<sub>3</sub> signalling. Nature 355: 446-449, 1992.
- 11. Marks MS, Hallenbeck TN, Nagata T, Segars JH, Appella E, Nikodem VM and Ozato K, H-2RIIBP (RXRB) heterodimerization provides a mechanism for combinatorial diversity in the regulation of retinoic acid and thyroid hormone responsive genes. EMBO J 11: 1419-1435, 1992.
- 12. Goodenough DA, Goliger JA and Paul DL, Connexins, connexons, and intercellular communication. Annu Rev Biochem 65: 475-502, 1996.
- 13. Hoffman BF and Rosen MR, Cellular mechanisms for cardiac arrhythmias. Circ Res 49: 1-15, 1981.
- 14. DeMello WC, Gap junctional communication in excitable tissues; the heart as a paradigma. Prog Biophys Mol Biol 61: 1-35, 1994.
- 15. Klein I, Thyroid hormone and the cardiovascular system. Am J Med 88: 631-637, 1990.
- 16. Stewart WW, Functional connections between cells as revealed by dye-coupling with a highly fluorescent naphthalimide tracer. Cell 14: 741-759, 1978.
- 17. Chomczynsci P and Sacchi N, Single-step method of RNA isolation by acid guanidinium thiocyanate-phenol-chloroform extraction. Anal Biochem 162: 156-159, 1987.
- 18. Hennemann H, Suchyna T, Lichtenberg-Frate H, Jungblut S, Dahl E, Schwarz J, Nicholson BJ and Willecke K, Molecular cloning and functional expression of mouse connexin40, a second gap junction gene preferentially expressed in lung. JCell Biol 117: 1299-1310, 1992.
- 19. Tso JY, Sun XH, Kao TH, Reece KS and Wu R, Isolation and characterization of rat and human glyceraldehyde-3-phosphate dehydrogenase cDNAs: genomic complexity and molecular evolution of the gene. Nucleic Acids Res 13: 2485-2502, 1985.
- 20. Oh SY, Grupen CG and Murray AW, Phorbol ester induces phosphorylation and down-regulation of connexin43 in WB cells. Biochim Biophys Acta 1094: 243-245, 1991.
- 21. Brümmer F, Zempel G, Bühle P, Stein JC and Hülser DF, Retinoic acid modulates gap junctional permeability: a comparative study of dye spreading and ionic coupling in cultured cells. Exp Cell Res 196: 158-163, 1991.
- 22. Hossain MZ and Bertram JS, Retinoids suppress proliferation, induce cell spreading, and up-regulate connexin43 expression only in postconfluent 10T1/2 cells: implications for the role of gap junctional communication. Cell Growth Differ 5: 1253-
- 23. Hanusch M, Stahl W, Schulz WA and Sies H, Induction of gap junctional communication by 4-oxoretinoic acid generated from its precursor canthaxanthin. Arch Biochem Biophys 317: 423-428, 1995.

- 24. Guo H, Acevedo P, Don Parsa F and Bertram JS, Gapjunctional protein connexin43 is expressed in dermis and epidermis of human skin: differential modulation by retinoids. *J Invest Dermatol* **99:** 460–467, 1992.
- Crow DS, Beyer EC, Paul DL, Kobe SS and Lau AF, Phosphorylation of connexin43 gap junction protein in uninfected and Rous sarcoma virus-transformed mammalian fibroblasts. Mol Cell Biol 10: 1754–1763, 1990.
- Rogers M, Berestecky JM, Hossain MZ, Guo H, Kadle R, Nicholson BJ and Bertram JS, Retinoic-enhanced gap junctional communication is achieved by increased levels of connexin43 mRNA and protein. Mol Carcinog 3: 335–343, 1990.
- 27. Bex V, Mercier T, Chaumontet C, Gaillard-Sachez I, Flechon

- B, Mazet F, Traub O and Martel P, Retinoic acid enhances connexin43 expression at the post-transcriptional level in rat liver epithelial cells. Cell Biochem Funct 13: 69-77, 1995.
- Clairmont A, Tessmann D and Sies H, Analysis of connexin43 gene expression induced by retinoic acid in F9 teratocarcinoma cells. FEBS Lett 397: 22–24, 1996.
- 29. Yu W, Dahl G and Werner R, The connexin43 gene is responsive to oestrogen. Proc R Soc Lond B Biol Sci 255: 125-132, 1994.
- Schräder M, Müller KM, Nayeri S, Kahlen J-P and Carlberg C, Vitamin D<sub>3</sub>-thyroid hormone receptor heterodimer polarity directs ligand sensitivity of transactivation. *Nature* 370: 382–386, 1994.